

Specific tools for genomics in UNIX: bedtools, bedops, vcftools,...

Course: Work with genomic data in the UNIX

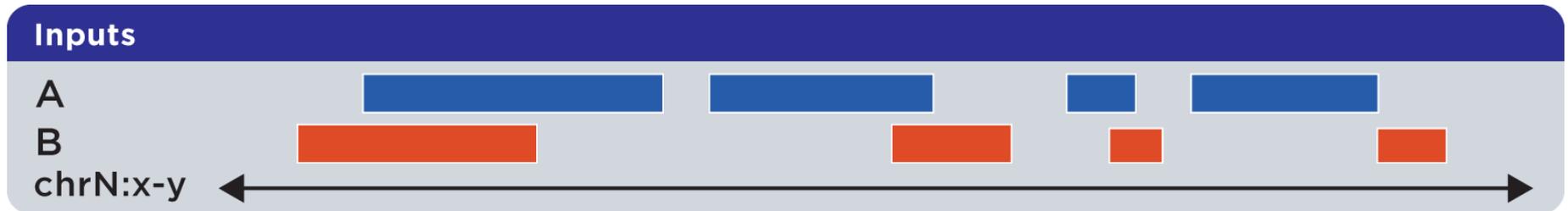
April 2015

Genome arithmetics

- Operations with genomic data based on their physical position in genome
- Variables:
 - chromosome
 - feature start, feature end
 - id
 - strand
- Basic data format: BED

Genome arithmetics: Examples

- Two sets of features (BED files):



<http://bedops.readthedocs.org>

```
chr1 1000 1200
chr1 1700 2100
chr2 1100 1500
```



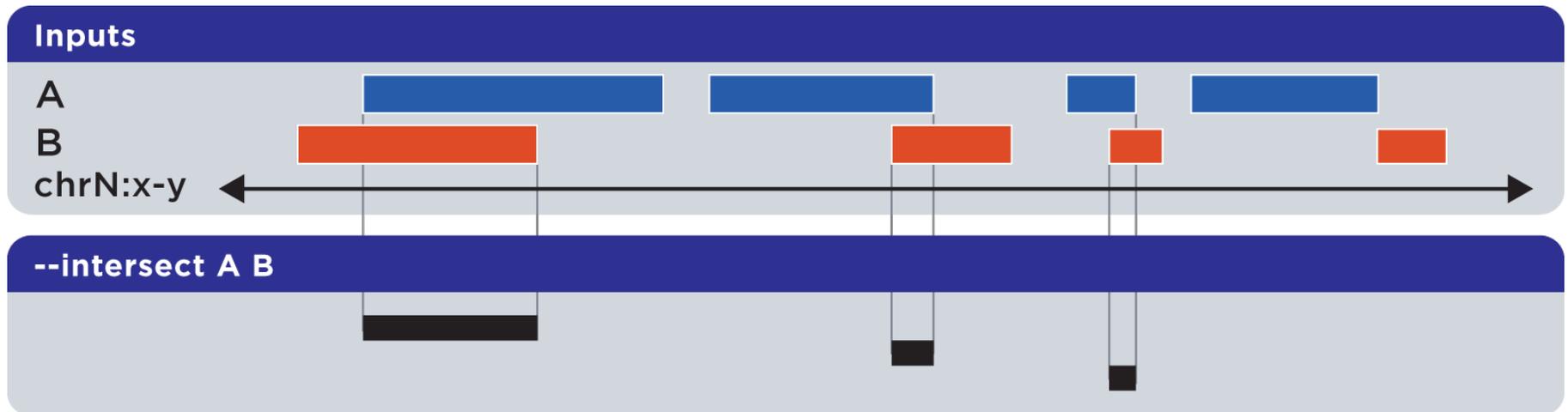
```
chr1 700 1100
chr1 1400 1500
chr1 1600 1900
```

New set of features based on combination of the previous sets using a specific rule

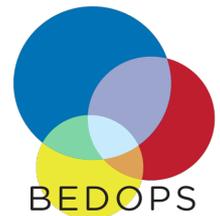


Genome arithmetics: Examples

- The rule: Get parts of features that overlap



<http://bedops.readthedocs.org>



```
1 9000 21000 gene1
1 30000 35000 gene2
1 65000 80000 gene3
2 32000 45000 gene4
2 55000 70000 gene5
```

```
1 8000 10000 feature1
1 16000 18000 feature2
1 24000 26000 feature3
1 38000 45000 feature4
1 60000 70000 feature5
2 10000 13000 feature6
2 40000 44000 feature7
```

```
bedops --intersect genes.bed features.bed
bedtools intersect -a genes.bed -b features.bed
```

```
1 9000 10000
1 16000 18000
1 65000 70000
2 40000 44000
```

Genome arithmetics: Examples

- The rule: Merge entire features



<http://bedops.readthedocs.org>



```
1 9000 21000 gene1
1 30000 35000 gene2
1 65000 80000 gene3
2 32000 45000 gene4
2 55000 70000 gene5
```

```
1 8000 10000 feature1
1 16000 18000 feature2
1 24000 26000 feature3
1 38000 45000 feature4
1 60000 70000 feature5
2 10000 13000 feature6
2 40000 44000 feature7
```

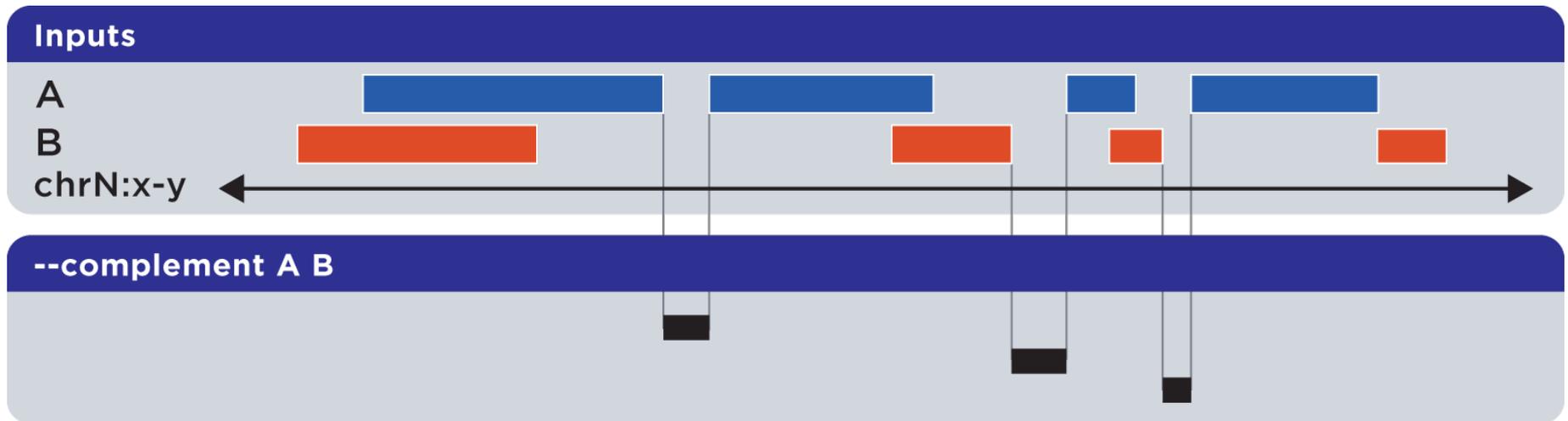
```
bedops --merge genes.bed features.bed
```

```
cat *.bed | sortBed > features2.bed
bedtools merge -i features2.bed
```

```
1 8000 21000
1 24000 26000
1 30000 35000
1 38000 45000
1 60000 80000
2 10000 13000
2 32000 45000
2 55000 70000
```

Genome arithmetics: Examples

- The rule: Get complement features



<http://bedops.readthedocs.org>



```
1 9000 21000 gene1
1 30000 35000 gene2
1 65000 80000 gene3
2 32000 45000 gene4
2 55000 70000 gene5
```

```
1 8000 10000 feature1
1 16000 18000 feature2
1 24000 26000 feature3
1 38000 45000 feature4
1 60000 70000 feature5
2 10000 13000 feature6
2 40000 44000 feature7
```

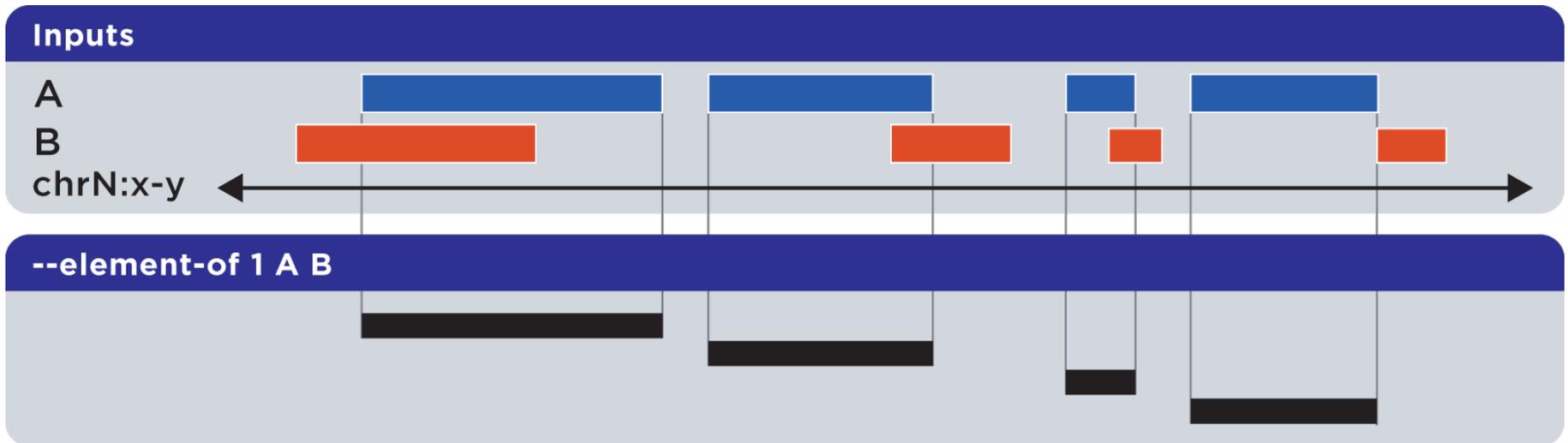
```
bedops --complement genes.bed features.bed
bedtools complement -i <(cat *.bed | sortBed) -g my.genome
```

```
1 21000 24000
1 26000 30000
1 35000 38000
1 45000 60000
2 13000 32000
2 45000 55000
```

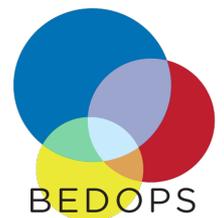
```
1 0 8000
1 21000 24000
1 26000 30000
1 35000 38000
1 45000 60000
1 80000 100000
2 0 10000
2 13000 32000
2 45000 55000
2 70000 120000
```

Genome arithmetics: Examples

- The rule: Report A which overlaps B



<http://bedops.readthedocs.org>



1	9000	21000	gene1
1	30000	35000	gene2
1	65000	80000	gene3
2	32000	45000	gene4
2	55000	70000	gene5

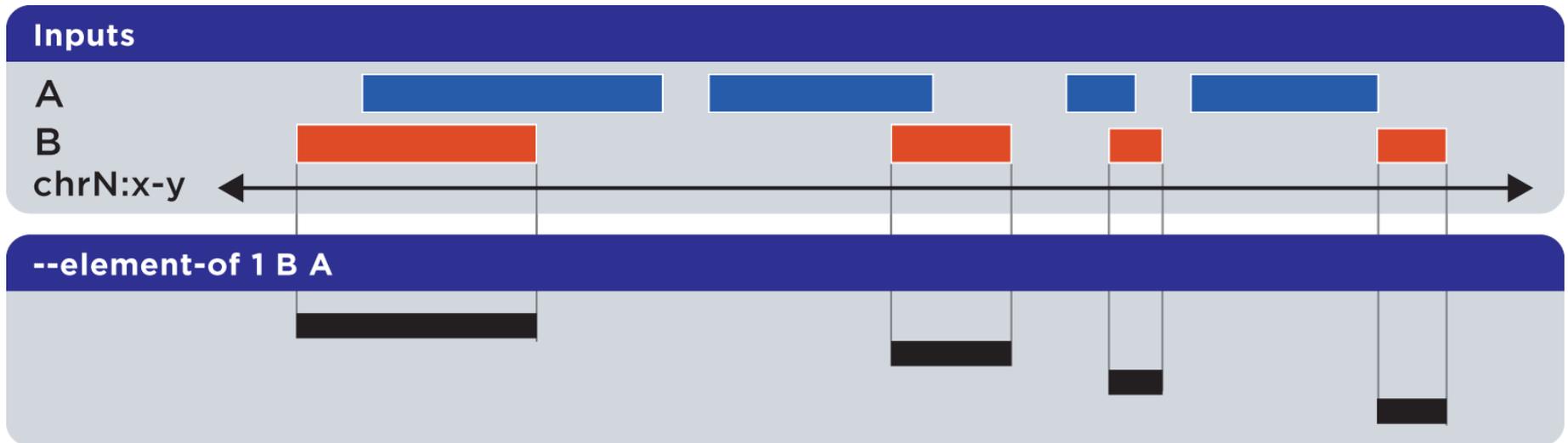
1	8000	10000	feature1
1	16000	18000	feature2
1	24000	26000	feature3
1	38000	45000	feature4
1	60000	70000	feature5
2	10000	13000	feature6
2	40000	44000	feature7

```
bedops --element-of 1 genes.bed features.bed  
bedtools intersect -u -a genes.bed -b features.bed
```

1	9000	21000	gene1
1	65000	80000	gene3
2	32000	45000	gene4

Genome arithmetics: Examples

- The rule: Report B which overlaps A



<http://bedops.readthedocs.org>



1	9000	21000	gene1
1	30000	35000	gene2
1	65000	80000	gene3
2	32000	45000	gene4
2	55000	70000	gene5

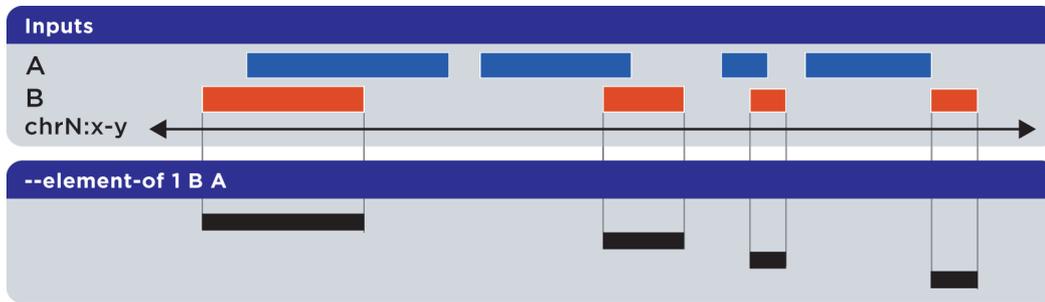
1	8000	10000	feature1
1	16000	18000	feature2
1	24000	26000	feature3
1	38000	45000	feature4
1	60000	70000	feature5
2	10000	13000	feature6
2	40000	44000	feature7

```
bedops --element-of 1 features.bed genes.bed  
bedtools intersect -u -a features.bed -b genes.bed
```

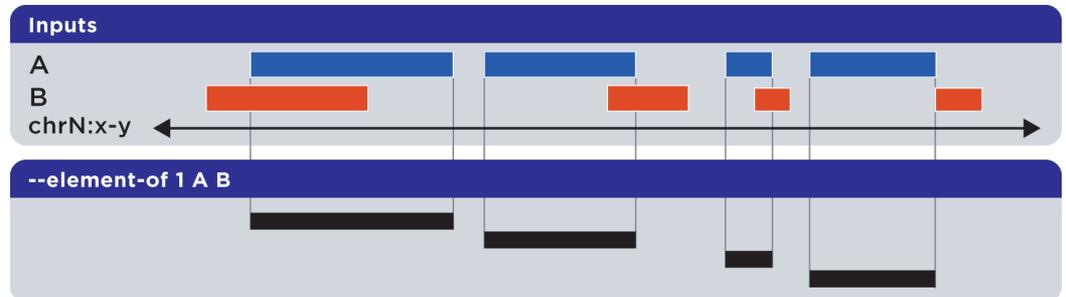
1	8000	10000	feature1
1	16000	18000	feature2
1	60000	70000	feature5
2	40000	44000	feature7

Genome arithmetics: Examples

- The rule: Report A,B which overlap each other



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<http://bedops.readthedocs.org>



1	9000	21000	gene1
1	30000	35000	gene2
1	65000	80000	gene3
2	32000	45000	gene4
2	55000	70000	gene5

1	8000	10000	feature1
1	16000	18000	feature2
1	24000	26000	feature3
1	38000	45000	feature4
1	60000	70000	feature5
2	10000	13000	feature6
2	40000	44000	feature7

```
bedtools intersect -wa -wb -a genes.bed -b features.bed
```

1	9000	21000	gene1	1	8000	10000	feature1
1	9000	21000	gene1	1	16000	18000	feature2
1	65000	80000	gene3	1	60000	70000	feature5
2	32000	45000	gene4	2	40000	44000	feature7

Genome feature summary

- Statistics, summary
- bedmap, bedtools (coverageBed, groupBy)
- e.g. depth coverage, base pair coverage, etc.

Genome feature summary: Example

- What is the base coverage of features within genes?

1	9000	21000	gene1
1	30000	35000	gene2
1	65000	80000	gene3
2	32000	45000	gene4
2	55000	70000	gene5

1	8000	10000	feature1
1	16000	18000	feature2
1	24000	26000	feature3
1	38000	45000	feature4
1	60000	70000	feature5
2	10000	13000	feature6
2	40000	44000	feature7

```

1 9000 21000 gene1
1 30000 35000 gene2
1 65000 80000 gene3
2 32000 45000 gene4
2 55000 70000 gene5

```

```

1 8000 10000 feature1
1 16000 18000 feature2
1 24000 26000 feature3
1 38000 45000 feature4
1 60000 70000 feature5
2 10000 13000 feature6
2 40000 44000 feature7

```

```

bedmap --echo --count --bases-uniq genes.bed features.bed
coverageBed -b genes.bed -a features.bed

```

```

1 9000 21000 gene1|2|3000
1 30000 35000 gene2|0|0
1 65000 80000 gene3|1|5000
2 32000 45000 gene4|1|4000
2 55000 70000 gene5|0|0

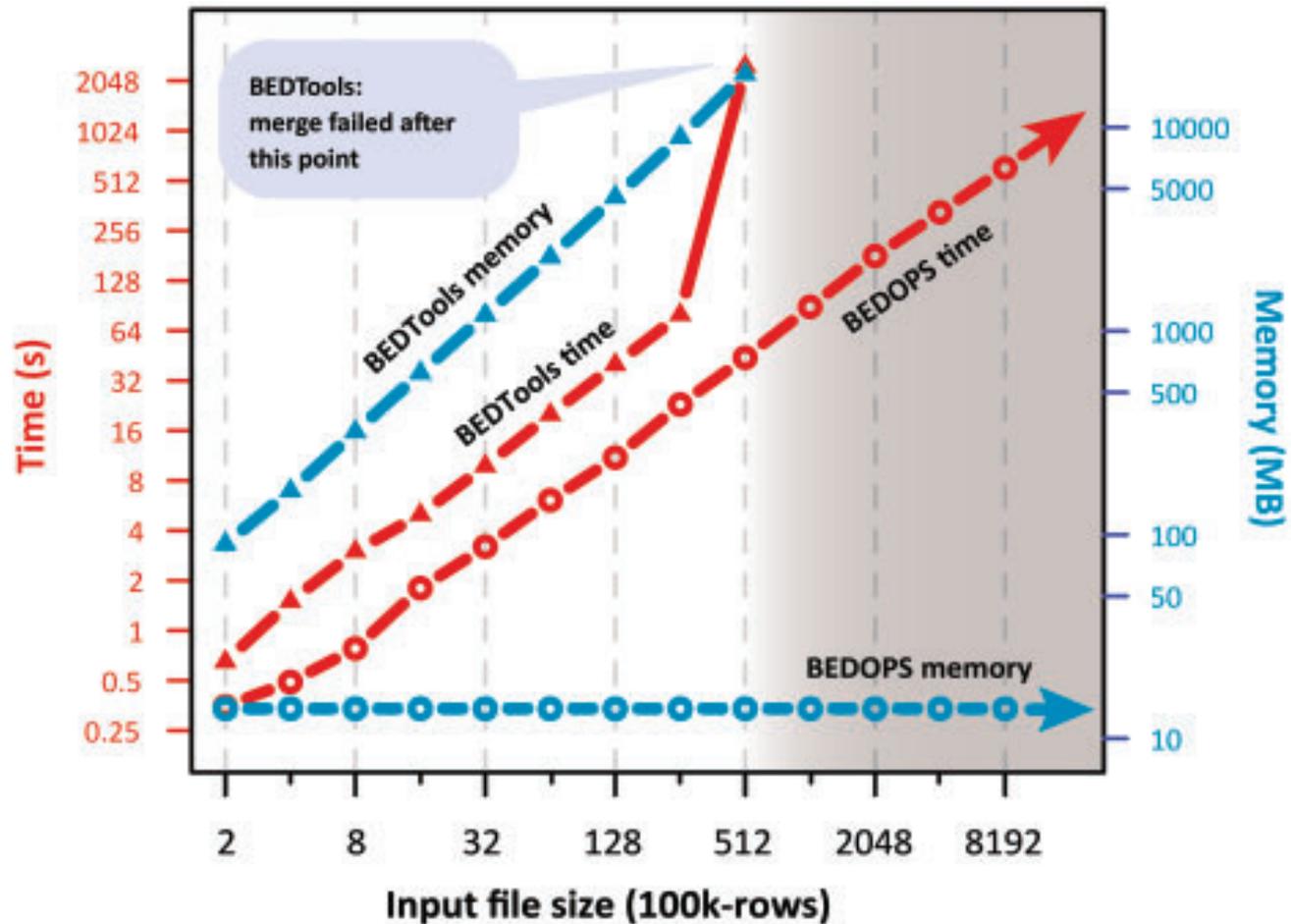
```

```

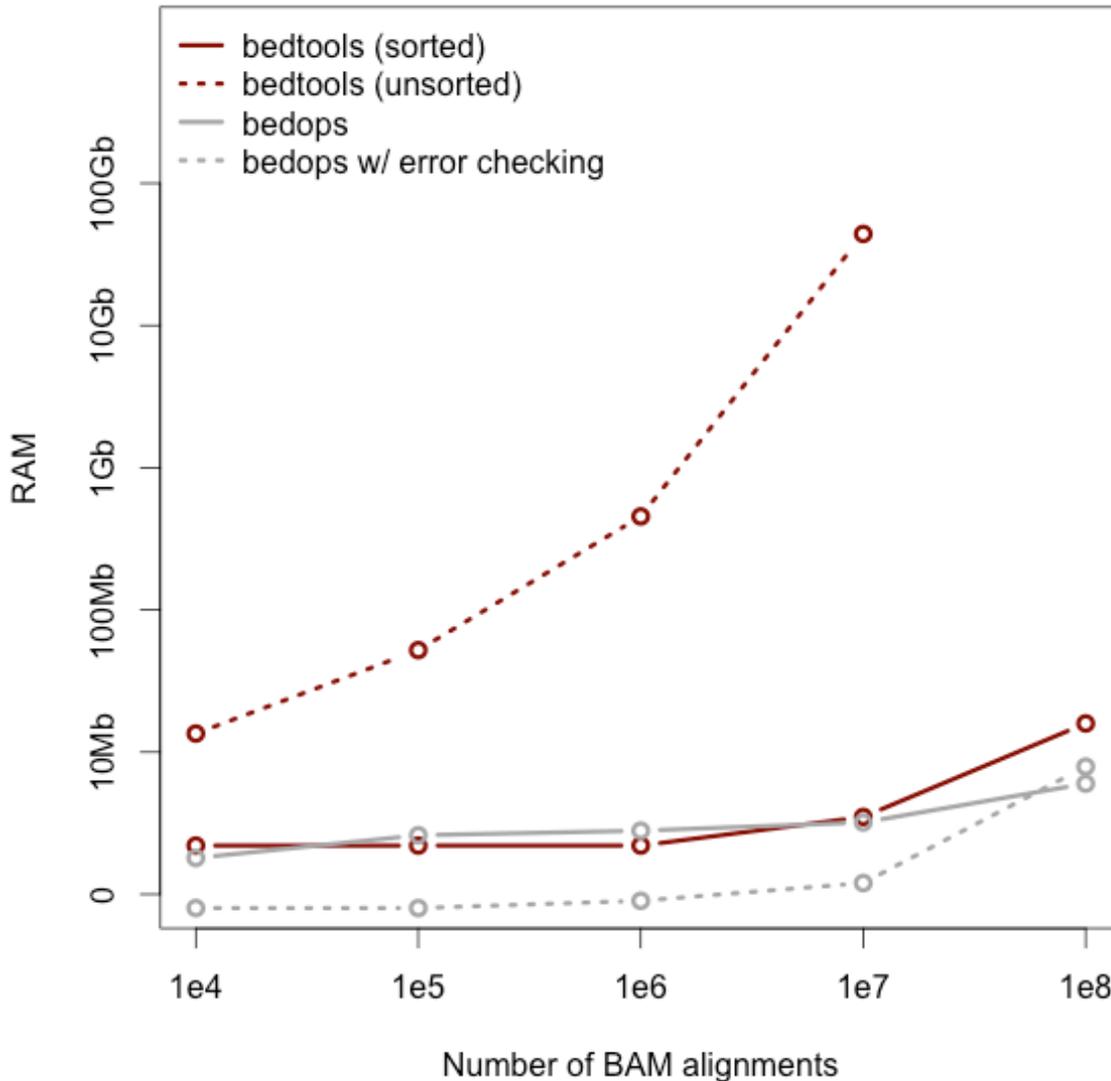
1 9000 21000 gene1 2 3000 12000 0.250000
1 30000 35000 gene2 0 0 5000 0.000000
1 65000 80000 gene3 1 5000 15000 0.3333333
2 32000 45000 gene4 1 4000 13000 0.3076923
2 55000 70000 gene5 0 0 15000 0.000000

```

bedtools vs. bedops



bedtools vs. bedops



Always use sorted data: sort-bed (bedops), sortBed (bedtools)

Other tools in bedtools

- makewindows
- cluster
- shuffle
- random
- jaccard
- reldist
- ...

Variation data: vcftools

- Efficient manipulation with VCF data
- Control quality
- Molecular evolution & population genetics measures/statistics
 - transition/transversion
 - heterozygosity, relatedness
 - Hardy-Weinberg
 - Weir & Cockerham's F_{st}
 - Nucleotide diversity
 - Linkage Disequilibrium

vcftools: starting

- Opening and viewing a vcf file:

```
vcftools --gzvcf popdata_mda.vcf.gz --recode --stdout | less -S
```

- Creating a new vcf file:

```
vcftools --gzvcf popdata_mda.vcf.gz --recode --out new_vcf
```

vcftools: data filtering

- Sample/Variant retrieval by name:
 - Individual/Variant names to keep/remove have to be specified in a separate file

```
--keep ind.txt # Keep these individuals
--remove ind.txt # Remove these individuals
--snps snps.txt # Keep these SNPs
--snps snps.txt --exclude # Remove these SNPs
```

```
vcftools --gzvcf popdata_mda.vcf.gz --keep euro_samples.txt --
recode --stdout | less -S
```

vcftools: data filtering

- Variant filtering based on physical location

```
--chr 11 # Keep just this chromosome
--not-chr 11 # Remove this chromosome
--not-chr 11 -not-chr 2 # Remove these two chromosomes
--from-bp 20000000 # Keep SNPs from this position
--to-bp 22000000 # Keep SNPs to this position
--bed keep.bed # Keep only SNPs overlapping with locations
listed in a file
--exclude-bed remove.bed # The opposite of the previous
```

```
vcftools --gzvcf popdata_mda.vcf.gz --keep euro_samples.txt --
chr 11 --from-bp 22000000 --to-bp 23000000 --recode --stdout |
less -S
```

vcftools: data filtering

- Variant filtering based on other features

```
--maf 0.2 # Keep just variants with Minor Allele Freq higher than 0.2
--hwe 0.05 # Keep just variants which do not deviate from HW equilibrium (p-value = 0.05)
--max-missing (0-1) # Remove SNPs with given proportion of missing data (0 = allowed completely missing, 1 = no missing data allowed)
--minQ 20 # Minimal quality allowed (Phred score)
```

```
vcftools --gzvcf popdata_mda.vcf.gz --keep euro_samples.txt --
recode --stdout | vcftools --vcf - --max-missing 1 -maf 0.2 --
recode --stdout | less -S
```

 stdin

vcftools: data filtering

- Variant filtering based on other features

```
--maf 0.2 # Keep just variants with Minor Allele Freq higher than 0.2  
--hwe 0.05 # Keep just variants which do not deviate from HW equilibrium (p-value = 0.05)  
--max-missing (0-1) # Remove SNPs with given proportion of missing data (0 = allowed completely missing, 1 = no missing data allowed)  
--minQ 20 # Minimal quality allowed (Phred score)
```

```
vcftools --gzvcf popdata_mda.vcf.gz --keep euro_samples.txt --  
recode --stdout | vcftools --vcf - --max-missing 1 -maf 0.2 --  
recode --stdout > popdata_mda_euro.vcf
```

vcftools: summary/statistics

- molecular evolution/population genetic

```
--site-pi # Calculates per-site nucleotide diversity ( $\pi$ )  
--window-pi 1000000 --window-pi-step 250000 # Calculates per-  
site nucleotide diversity for windows of 1Mb with 250Kb step  
--weir-fst-pop pop1.txt --weir-fst-pop pop2.txt # Calculates  
Weir & Cockerham's Fst  
--fst-window-size 1000000 --fst-window-step 250000 #  
Calculates Fst for windows of 1Mb with 250Kb step
```

```
vcftools --vcf popdata_mda_euro.vcf  
--weir-fst-pop musculus_samps.txt  
--weir-fst-pop domesticus_samps.txt --stdout | less -S
```

Exercise: Population differentiation

- Get a population differentiation calculated as F_{st} between *M. m. musculus* and *M. m. domesticus* within a given sliding window and find candidate genes within highly differentiated regions
 - use vcftools to filter data and calculate F_{st} for individual SNPs
 - use bedtools makewindows to create sliding windows of three sizes
 - 100 kb + 10 kb step
 - 500 kb + 50 kb step
 - 1 Mb + 100 kb step
 - use bedmap (bedops) to calculate average F_{st} for each window
 - use Rstudio and ggplot2 to plot F_{st} values across the genome
 - use R to obtain 99th percentile and use it to obtain a set of candidate genomic regions
 - use bedtools intersect to get a list of candidate genes

Exercise: Population differentiation

1. use vcftools to filter data and calculate Fst for individual SNPs

```
## Prepare files
```

```
cd
```

```
mkdir data/diff
```

```
cp /data/mus_mda/00-popdata/*.txt data/diff/.
```

```
mv /data/mus_mda/00-popdata/popdata_mda.vcf.gz data/diff/.
```

```
cd data/diff/
```

```
vcftools --gzvcf popdata_mda.vcf.gz --keep euro_samples.txt --  
recode --stdout | vcftools --vcf - --max-missing 1 -maf 0.2 --  
recode --stdout > popdata_mda_euro.vcf
```

Exercise: Population differentiation

1. use vcftools to filter data and calculate Fst for individual SNPs

```
vcftools --gzvcf popdata_mda.vcf.gz --keep euro_samples.txt --
recode --stdout | vcftools --vcf - --max-missing 1 -maf 0.2 --
recode --stdout > popdata_mda_euro.vcf
```

```
vcftools --vcf popdata_mda_euro.vcf
--weir-fst-pop musculus_samps.txt
--weir-fst-pop domesticus_samps.txt --stdout |
tail -n +2 |
awk -F $'\t' 'BEGIN{OFS=FS}{ print $1,$2-1,$2,$1":"$2,$3}' >
popdata_mda_euro_fst.bed
```

Exercise: Population differentiation

2. use bedtools makewindows to create sliding windows of three sizes
 - 100 kb + 10 kb step
 - 500 kb + 50 kb step
 - 1 Mb + 100 kb step

Inputting from subshell
<(command producing input)

```
cp /data/mus_mda/02-windows/genome.fa.fai .
```

```
bedtools makewindows -g <(grep '^2\|^11' genome.fa.fai) -w  
1000000 -s 100000 -i winnum | awk '{ print $0":1000kb" }' >  
windows_1000kb.bed
```

```
cat windows_*.bed > windows.bed
```

Exercise: Population differentiation

3. use bedmap (bedops) to calculate average Fst for each window

```
sort-bed windows.bed > windows_sorted.bed
sort-bed popdata_mda_euro_fst.bed >
popdata_mda_euro_fst_sorted.bed

bedmap --echo --mean --count windows_sorted.bed
popdata_mda_euro_fst_sorted.bed | grep -v NA |
tr "|:" "\t" > windows2snps_fst.bed
```

Exercise: Population differentiation

4. use Rstudio and ggplot2 to plot Fst values across the genome

```
library(ggplot2)

setwd("~/data/diff")

fst <- read.table("windows2snps_fst.bed", header=F, sep="\t")

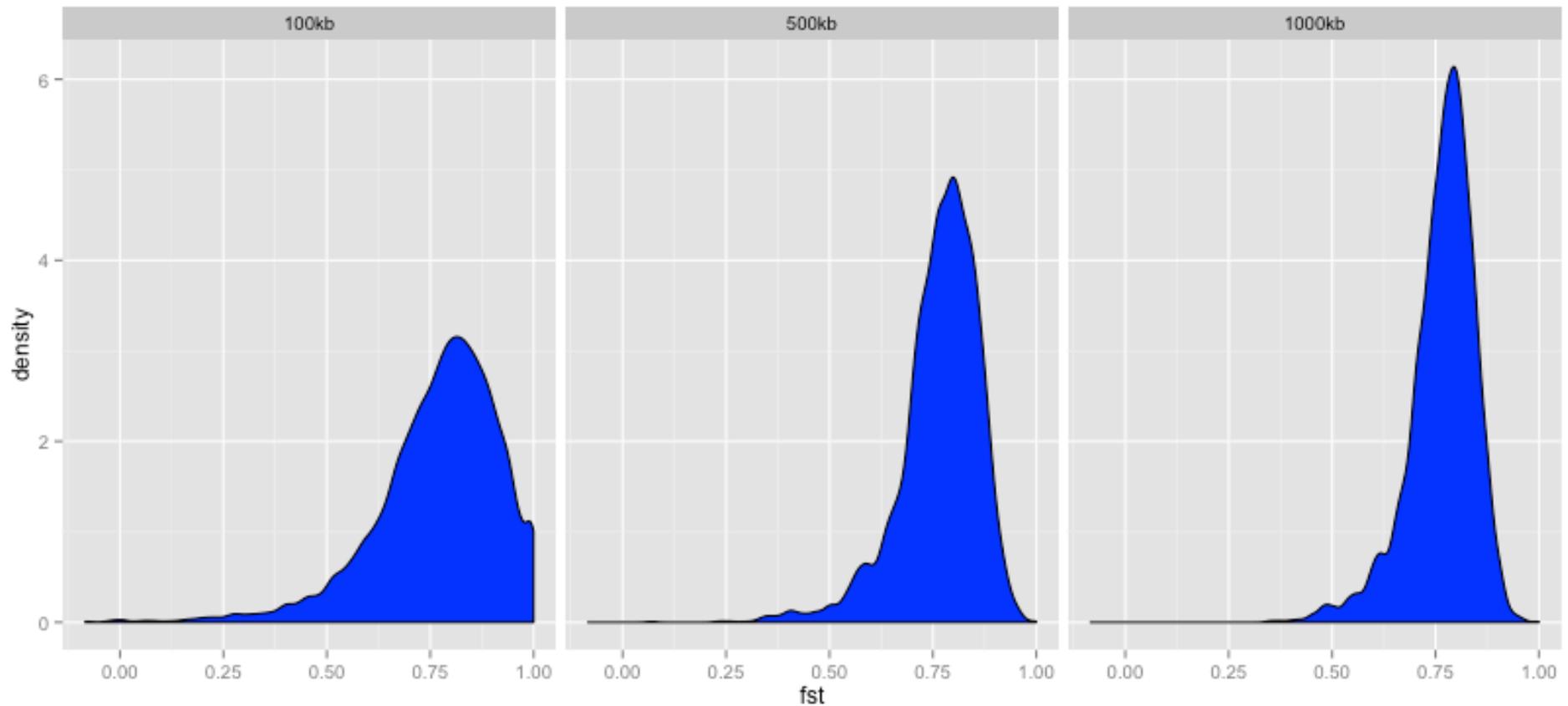
names(fst) <- c("chrom", "start", "end", "win_id", "win_size",
               "fst", "cnt_snps")

fst$win_size <- factor(fst$win_size, levels=c("100kb",
                                              "500kb", "1000kb"))

ggplot(fst, data=fst, geom="density", fill=I("blue")) +
  facet_wrap(~win_size)
```

Exercise: Population differentiation

4. use Rstudio and ggplot2 to plot Fst values across the genome



Exercise: Population differentiation

4. use Rstudio and ggplot2 to plot Fst values across the genome

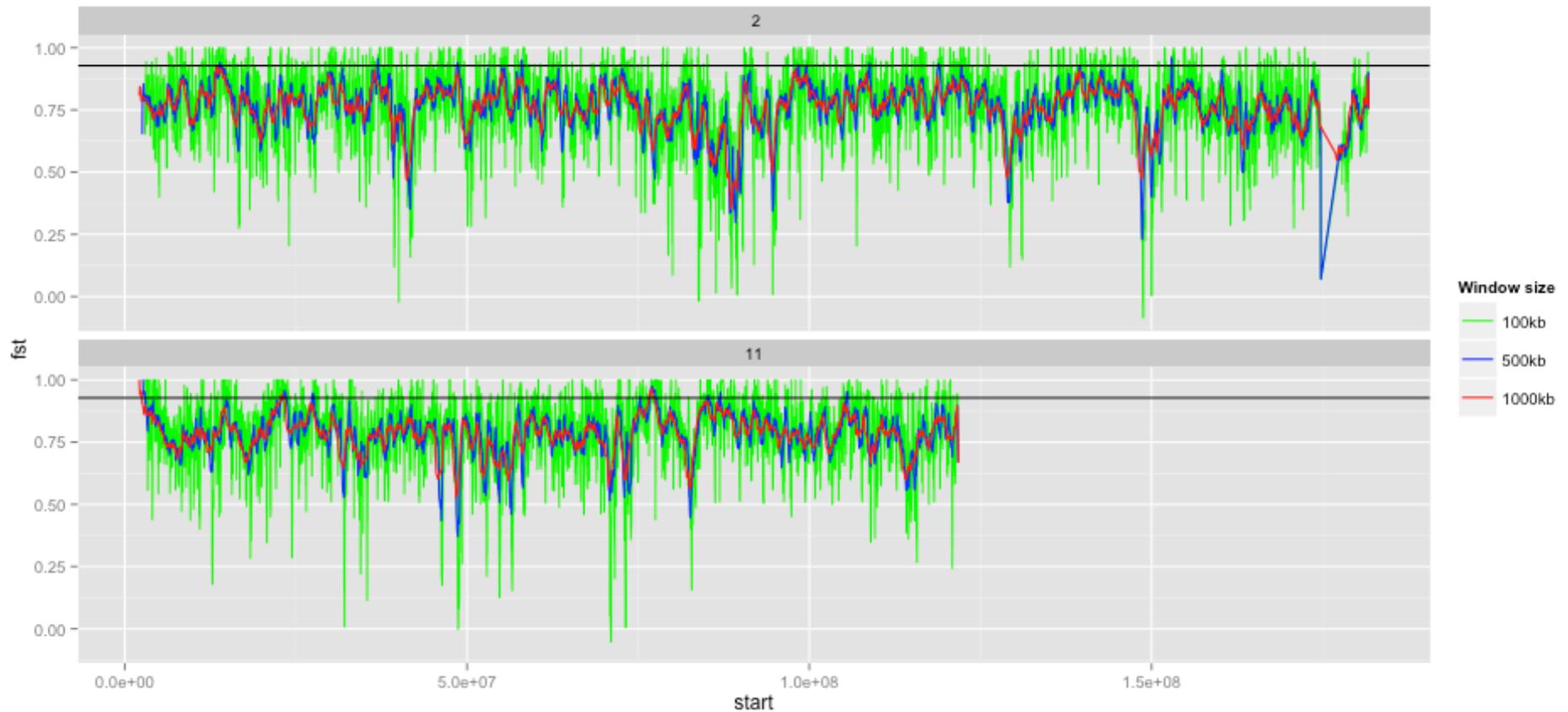
```
ggplot(fst, aes(y=fst, x=start, colour=win_size)) +  
  geom_line() +  
  facet_wrap(~chrom, nrow=2) +  
  scale_colour_manual(name="Window size", values=c("green",  
"blue", "red"))
```

```
q <- quantile(subset(fst, win_size=="500kb", select="fst")[,  
1], prob=0.99) [[1]]
```

```
ggplot(fst, aes(y=fst, x=start, colour=win_size)) +  
  geom_line() +  
  facet_wrap(~chrom, nrow=2) +  
  geom_hline(yintercept=q, colout="black") +  
  scale_colour_manual(name="Window size", values=c("green",  
"blue", "red"))
```

Exercise: Population differentiation

4. use Rstudio and ggplot2 to plot Fst values across the genome



Exercise: Population differentiation

5. use R to obtain 99th percentile and use it to obtain a set of candidate genomic regions

Use of Shell variables

var=value

var=`command`

echo \$var

```
q500=`grep 500kb windows2snps_fst.bed | cut -f 6 | Rscript -e  
'quantile(as.numeric(readLines("stdin")),p=c(0.99))[[1]]' |  
cut -d " " -f 2`
```

```
echo $q500
```

```
grep 500kb windows2snps_fst.bed | awk -v a=$q500 -F $'\t'  
'BEGIN{OFS=FS}{ if($6 >= a){print $1,$2,$3} }' |  
bedtools merge -i stdin > signif_500kb.bed
```

Exercise: Population differentiation

6. use bedtools intersect to get a list of candidate genes

```
bedtools intersect -a signif.bed -b  
Mus_musculus.NCBIM37.67.gtf -wa -wb | grep protein_coding |  
cut -f 1,2,3,4,13 | cut -d ' ' -f 1,3,9 | tr -d '";' | sort |  
uniq > fst2genes.tab
```